

by Burkard traps, HiVol traps, Rotocorols, or personal sampling must be sufficiently high to fulfill the other Thomsen postulates. Annual pollen sampling data in various regions of the country are available at the National Allergy Bureau web site (www.naab.org).

Aerobiologic variability also affects pollen distribution. Misting, the simultaneous production of large numbers of pollens by a plant population, is a common feature among trees in temperate forests.²⁶ Allergenicity is enhanced with higher daily mean temperatures.²⁷ Successive wetting and drying cycles release not only pollen but also very small cytoplasmic fragments (30 nm to 4 μ m) that retain allergenic activity.^{28,29} The latter are detected by immunostaining of personal cascade impactors.^{30–32} Pollen distribution also varies with altitude.^{33–36}

The difference between cross-sensitization and cross-sensitization is often misunderstood in the selection of relevant skin test and extract reagents. The Allergome database revealed that pollen allergens can be classified into 29 of 7,868 protein families.³⁷ Parallegens such as profilin, polcalicin, l.3. β -glucanase, and cross-reactive carbohydrate determinants account for extensive cross-reactivity among pollen-sensitized patients.^{38–41} Thus, the relevant allergen profile of ash shares epitopes with pollen allergens not only from other tree pollens but also from grass and weed pollen species.⁴² Similarly, pollen allergy to white birch, a member of the order Fagales, can be found in birch-free geographic areas that have other non-birch tree species of the Fagales order.⁴³ This is particularly germane to the decision about number of tests because it demonstrates clearly that skin test reactivity alone cannot decide the clinical significance of an allergen. A high percentage of reactions to the ornamental black locust in pollinosis patients is ascribed to cross-sensitization to parallegens in other common pollens.⁴⁴ This is termed an allergy surrogate.⁴⁵ Cross-sensitivity to pollen profiles has been demonstrated in CD4⁺ T_H2 clones, which promiscuously recognize homologically conserved regions on birch and grass profiles.⁴⁶ This may be in part due to conserved allergen-specific motifs.⁴⁷ Cross-sensitization to profilin and/or bromelain-type cross-reactive carbohydrate determinants caused by timothy grass or ragweed pollen has also been reported in venom sensitization.^{48,49} These recent demonstrations of extensive cross-sensitivity among all pollens must be critically reviewed either when selecting a skin test panel or when interpreting the results.

The reported prevalence of outdoor airborne fungi depends on sampling technique (viable vs nonviable; bioaerosol vs surface) and the collecting device.^{50–52} Viable cultures also vary depending on media and duration of culture. In general, *Cladosporium* and *Aspergillus* species are predominant in the summer months. Indoor mold sampling almost always detects species of *Aspergillus* and *Penicillium*.^{53–56} Recent molecular cloning of airborne fungal spores can also identify many *Bovine*mycelium and *Ascomycetes*.⁵⁷ Although many other species have been identified, their comparative significance is

difficult to ascertain. Cross-allergenicity among major classes of airborne fungi has not been well delineated.

These facts about cross-allergenicity are particularly germane to formulation of treatment extracts for a particular floristic region, which should reflect the validity of aerobiologic sampling, the constitutive allergenicity of pollens, as evaluated by direct skin exposure, and/or challenge tests, and how well positive test results correlate with the patient's clinical symptoms. Other factors that may need to be incorporated into the final formulation decision include unexpected allergen exposure because of frequent travel to other floristic zones and commercial availability of appropriate allergen extracts.

Food prick and puncture skin tests are excellent diagnostic modalities for the diagnosis of IgE-mediated clinical entities, which include anaphylaxis, food-dependent exercise-induced anaphylaxis, acute urticaria, atopic dermatitis, and the oral allergy syndrome. The last is often associated with cross-sensitivity to parallegens in pollens.⁵⁸ In many instances, the history suggests appropriate allergen testing; in other situations, a preliminary diet history and diaries provide additional clues. At times, reconstruction of a suspected etiologic meal may direct suspicion to specific food components in that meal. Relatively few foods account for most IgE-mediated allergic reactions in both children and adults. The more common food allergens in infants and young children are cow's milk, hen's egg, peanuts, tree nuts, soybeans, and wheat, whereas the adult counterparts are peanuts, tree nuts, fish, crustaceans, mollusks, fruits, and vegetables. Commercial fruit and vegetable extracts rapidly lose potency so many clinicians either prepare fresh extracts of these classes of foods or test by the prick-prick method. This method is preferred to detect strain differences in fruit allergens (eg, apple).⁵⁹ Because of many false-positive test results and potential risks, intracutaneous tests to foods are not recommended. Food tests are inappropriate for investigation of chronic idiopathic urticaria (CIU) or angioedema.

The scope and number of skin tests for allergy diagnosis reflect the clinician's scientific knowledge and clinical experience. The choice and number of test allergens should be continuously refined in accord with scientific advances, botanic and aerobiologic surveys, demographic trends, and availability of relevant, defined reagents. Practice must be directed to the best documented concepts of allergen prevalence, geographic distribution, and immunochemical relationships.

Although no prospective studies provide direct evidence for these issues, the literature concerning clinically relevant allergens suggests a rationale and evidence-based process for determining the number of skin test reagents. This issue had received serious consideration by the Joint Task Force on Practice Parameters in conjunction with expert consultants during the preparation of Allergen Immunotherapy: A Practice Parameter, at which time the Joint Task Force suggested a core panel of indoor and outdoor inhalant allergens. This list includes representative species of the major classes of

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JS Bruner



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