#### HIGH-PERFORMANCE THIN LAYER CHROMATOGRAPHY DETERMINATION OF ASTAXANTHIN IN Emphassic superbo OIL.

#### Meng Huang and Duicheng Liu\*

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The Antarctic krill (Expérience asperèn) is a kind of marine cooplankton belonging among Crustacca [1]. It is the principal prey of many marine pseudons by reason of its gigantic population [2]. The study of Antarctic krill has become a hot research field in the past few decades. It is not only the key organism in the southern occan but also the biggest fishery resource in the world. With its high content of astaxamilin as a valuable natural pseudoct [2, 3], it urgently awaits development and utilization. The ketocaroteneid astaxamilin  $(5.3^{\circ}\text{--dibydrosy-}\beta_i\beta^i\text{--carotene-}4.4^{\circ}\text{--dione})$  and astaxamilin esters are the principal components of Antarctic krill pigment [4], and astaxamilin is one of most important carotenoids by virtue of its biologic functions as a vitamin A processor and as a much more efficient anticoldant than  $\beta$ -carotene and vitamin E [5]. It is also a natural coloring (in fish muscles, bird feathers, yofks, etc.), supports the immune system, has an anti-turnor effect, and protects from ultraviolet indication [6–8]. Overall, astaxamilin can play an important role in healthcare and connectic manufacture, with enormous commercial and industrial prespects:

As a general rule, high-performance liquid chromatography (HPLC) is preferred in astaxanthin assay due to its wide use all around the world [9], whereas the HPTLC technique with high sensitivity and resolution can also function excellently in the separation and parification of bioactive compounds from natural materials.

In short, we aim to establish and validate as HPTLC method that is adapted to astaxanthin determination in Antarctic krill oil and that can be applied to other materials that contain astaxanthin.

The astoxanthin in Antarctic krill is mostly in the form of astoxanthin esters for its molecular stabilization [10]. Some are diesters and others are monoesters, attributed to the two hydroxyls that can combine with different fitty acids at both ends of the astoxanthin molecule [11]. While the concentration of free astoxanthin was small, all of it and other impurities are fat soluble and can be dissolved in low-polarity reagents; thus, whexare-acetone (7:2, w/v) was chosen to be the mobile phase after repeated attempts, combined with the high-performance silica gel 60 F254, which can accomplish the HPTLC separation within 10 min. The R<sub>2</sub> value of the free astoxanthin was 0.32.

It was necessary to apply the suponification procedure to release the astaxanthis combined with fatty acids because the concentration of free astaxanthis was from  $5\pm1.0\%$  to  $85\pm5.0\%$ , so that the qualification and quantification of estaxanthis can be carried out more successfully, as well as the further purification, since there was no standard for astaxanthin esters. KORL was essential in suponification, but it can result in significant degradation of astaxanthin, especially when the reaction temperature was high. There was almost no degradation of astaxanthin at  $4^{\circ}C$  [12]; therefore it was selected as the suponification temperature, and the maximum concentration of free astaxanthin can reach 89.7466%. The linear regression equation of astaxanthin after suponification is Y = 1309.2935 + 47.9693 X (where Y is the response and X is the mass of astaxanthin). The correlation coefficient was found to be 0.9990, the RSD (relative standard deviation) was 2.56%, and the astaxanthin in the suponification sample was 0.7445 mg mit. 1: more data are shown in Table 1.

The three main components in the original sample were separated without interference from other impurities, and the spectral evaluation of the samples showed the same maximum absorption ( $\lambda_{ress}$  476 nm) as the standard; therefore, the specificity of this method is verified.

The RSD of introday precision was 1.68% and the value of the interday precision was 5.07%. Both values were determined when the plates were stored in the dark.

The accuracy was clarified as percent recovery, and the average recovery of astaxanthin was 98,53%.

Key Laboratory of Animal Resistance, College of Life Science, Shandong Normal University, 88 East Wenhua Road, 250014, Jimm, P. R. China, fix: 86 0531 86185360, e-mail: lindchii sdru.edu.ca. Published in Khimon Prirochykh Socchwenii, No. 1, January February, 2013, pp. 128–129. Original article submitted September 23, 2011.

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Eli Grushka, Nelu Grinberg

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